**In vitro** production of gases with mixtures of *Hyparrhenia rufa* (Nees) and *Leucaena leucocephala* (Lam) de Wit

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**ABSTRACT**

**Objective**: To evaluate total **in vitro** gas and methane (CH₄) production in different mixtures of *Hyparrhenia rufa* (Hr) and *Leucaena leucocephala* (Ll).

**Design/methodology/approach**: In airtight biodigesters with 200 mL of culture medium, 20 g of the following treatments were incubated by triplicate: T1: 100% Hr, T2: 80% Hr + 20% Ll, T3: 60% Hr + 40% Ll, T4: 40% Hr + 60% Ll; these were inoculated with 20 mL of fresh rumen fluid and incubated at 38 ± 0.5 °C for 24, 48, 72 and 96 h. The total gas and CH₄ production were assessed; the data were analyzed in a completely randomized design.

**Results**: The addition of 20%, 40% and 60% Ll in mixture with Hr decreased the neutral detergent fiber (NDF), acid detergent fiber (ADF), total gas and CH₄ production, while the crude protein content increased.

**Study limitations/implications**: In vivo studies are required / by including amounts higher than 20% Ll may improve energy utilization efficiency.

**Findings/conclusions**: Adding more than 20% *L. leucocephala* in a mixture with *H. rufa* decreases total gas and CH₄ production.

**Keywords**: methane, ruminants, tropical grasses, forage shrubs.

**INTRODUCTION**

The digestive process of the food consumed by ruminants involves physical, microbiological and chemical processes. The fermentation of food in the rumen is carried out by microorganisms, including bacteria, protozoa and fungi; the products of fermentation are: energy in the form of adenine triphosphate (ATP), which is used by microorganisms to grow and reproduce, the process involves the synthesis of microbial mass that will be later digested in the abomasum and used by the ruminant as a true protein source; volatile fatty acids such as acetic, propionic and butyric are also produced, which will be used by the ruminant as a principal energy source. As secondary products of fermentation,
carbon dioxide (CO₂), hydrogen (H₂) and CH₄ are produced, these are synthesized from the fermentation of structural carbohydrates products by rumen methanogenic archaea such as Methanobrevibacter ruminantium, Methanobacterium formicicum, Methanomicrobium mobile (Cobos et al., 2018).

The CH₄ production is necessary for the oxidation of nicotin adenine dinucleotide (NAD), it is a step in the process for obtaining the energy contained in the neutral detergent fiber (NDF) and other nutrients in the diet, due that this process reduces the accumulation of H₂, it regulates pH and reduces ruminal pressure (Sharp et al., 1998; Ley de Coss et al., 2018) to maintain ruminal stability (Galindo et al., 2010). In a second process, with the manure fermentation, nitrous oxide (N₂O) is produced, a gas with 310 times the potential of heat retaining than CO₂ (Ellis et al., 2012).

Methane emissions from bovine enteric fermentation account for about 39% of the greenhouse gases (GHG) produced by the livestock sector (Arbre et al., 2016), it is necessary to reverse the negative trends of extensive livestock farming, where the increase of CO₂ was of 70% and 40% for CH₄ during the 1970 to 2004 period (IPCC, 2016). To achieve this, it is required to design and evaluate methodologies to accurately calculate the energy flow and its relationship with GHG emissions in livestock activity (De-Vries and de-Boer, 2010).

In the southern and southeastern regions of Mexico, the ruminants’ diet is based on forage grasses and shrubs; the former has a higher NDF, acid detergent fiber (ADF) and lower protein content, which favors higher CO₂, H₂ and CH₄ production. Methane production in tropical regions is of 20 L kg⁻¹ of fermented dry matter (Piñeiro et al., 2017), with energy losses in the form of CH₄ of up to 18%, so that, by using feeding strategies that allow greater efficiency in energy utilization, emissions of this gas could be reduced and cattle productivity improved (Vanlierde et al., 2016).

The rate of CH₄ emissions from ruminal fermentation is directly related to the intake, nutrient content, physical and chemical characteristics and diet digestibility. Therefore, a strategy to reduce enteric methane production is through feeding management that allows changes in the ruminal fermentation pattern (Piñeiro et al., 2017). The inclusion of forage trees and shrubs in the feeding of ruminants has improved NDF degradation, due to the higher crude protein contribution. When 20% of L. leucocephala was included in the diet, CH₄ production was reduced by 26% (Piñeiro et al., 2015). Reduction in CH₄ emission was also observed with the shrubs as L. leucocephala, Sapindus saponaria, Calliandra calothyrsus, Pithocellobium dulce, Helicarpus velutinus and Guazuma ulmifolia, when supplied with low quality pasture or with Megathyrsus maximus (Gaviria et al., 2015; Lopez et al., 2016). Based on the above, the objective was to assess the total gases and CH₄ production, when combined with Hyparrhenia ruffa grass and Leucaena leucocephala in in vitro incubation.

MATERIALS AND METHODS

Location of the study. The research took place from May 1 to June 30, 2019, in the Jaulas Metabólicas area and the Laboratorio de Sanidad Agropecuaria at the Facultad de Ciencias Agronómicas, Campus V of the Universidad Autónoma de Chiapas located at the Centro Universitario de Transferencia de Tecnología (University Center for Technology Transfer, CUTT) “San Ramón”, Carretera al ejido 16 de septiembre km 2.5 in Villaflor,
Chiapas, Mexico (16° 27’ 59” N and 93° 28’ 43” W). From September 2 to 16, 2019, the analyses were performed at the Laboratorio de Nutrición Animal of the Programa en Ganadería from the Colegio de Postgraduados, Campus Montecillos, México (19° 27’ 39.24” N and 98° 54’ 29.19” W).

**Chemical analysis of samples.** The mixtures of *H. rufa* and *L. leucocephala* were analyzed for total dry matter (TDM), crude protein (CP) following the Kjeldahl method, ethereal extract (EE) via the Soxhlet method, crude fiber (CF) by the Weende method, ash (C) and organic matter (OM) by difference, all using the techniques described by the AOAC (2012). The fiber fractions (NDF and FDA) determination was performed with alpha-amylase without ash correction as specified by Van Soest *et al.* (1991).

**Growth medium and treatments.** Rumen liquid (RL) from a 525 kg live weight male Brown Swiss bovine was used, the RL was extracted with an esophageal tube with a vacuum pump. The bovine received a diet with 85% *Hyparrhenia rufa* and 15% of a concentrated feed, containing 2.7 Mcal of ME and 14% crude protein (CP), the culture medium used to determine total gas (TG) production, CO2 and CH4 ratios and DM degradation (Table 1), was prepared in sterile conditions and low CO2 flow. The evaluated treatments were: T1: 100% *Hyparrhenia rufa* (Hr), T2: 80% Hr + 20% *L. leucocephala* (Ll), T3: 60% Hr + 40% Ll, T4: 40% Hr + 60% Ll.

**Table 1.** Culture medium for *in vitro* fermentation of dry matter of *Hyparrhenia rufa* and *Leucaena leucocephala* mixtures.

<table>
<thead>
<tr>
<th>Compound</th>
<th>Quantity (mL) for 100 mL of medium</th>
</tr>
</thead>
<tbody>
<tr>
<td>Destilled water</td>
<td>52.9</td>
</tr>
<tr>
<td>Clarified rumen fluid</td>
<td>30.0</td>
</tr>
<tr>
<td>Mineral solution I</td>
<td>5.0</td>
</tr>
<tr>
<td>Mineral solution II</td>
<td>5.0</td>
</tr>
<tr>
<td>Sodium carbonate (Na2CO3), 8%</td>
<td>5.0</td>
</tr>
<tr>
<td>Sulfide-cysteine solution</td>
<td>2.0</td>
</tr>
<tr>
<td>Resazurin solution, 0.1 %</td>
<td>0.1</td>
</tr>
</tbody>
</table>

(1) Clarified rumen fluid filtrated and centrifugated at 17,664 g for 15 min and sterilized 20 min at 21 °C a 15 psi. (2) Contains (in 1000 mL) 6 g K2HPO4. (3) Contains (in 1000 mL H2O), 6 g KH2PO4, 6 g (NH4)2SO4, 12 g NaCl, 2.45 g MgSO4 y 1.6 g CaCl2·H2O. (4) 8 g Na2CO3 in 100 mL H2O destilled. (5) 2.5 g L-cysteine (in 15 mL 2N NaOH) + 2.5 g Na2S·9H2O (in 100 mL H2O). (6) 0.1 mL resazurin in 100 mL.

**Production of total gases and CH4.** The *in vitro* production of TG and CH4 was determined by triplicate with repetition throughout time using 2.0 L capacity biodigesters with a hermetic look, to which the following mixture was added in aseptic conditions and CO2 flow: 20 g of DM of each treatment plus 200 mL of culture medium (Table 1), each treatment was inoculated with 20 mL of fresh RL, which was filtered in cotton gauze and kept in a water bath at 38±0.5 °C and CO2 flow. The initial total bacteria concentration was counted with the most probable number (MPN) technique with $1.30\times10^8$ CFU mL$^{-1}$.
at pH 6.72. The inoculated treatments were incubated for 24, 48, 72 and 96 h in a water bath at 38±0.5 °C. At the end of the incubation period, the TG pressure was measured with a manometer adapted to the equipment. Afterward, the amount of produced TG in the system was assessed using the liquid displacement technique through a trap with Mariotte flasks.

The displaced water was collected in a graduated cylinder and the amount of gas produced per gram of fermented DM (mL of gas g⁻¹ of MSf) was determined. In a second run and under the same culture conditions and temperature, a 2N NaOH solution (20 g L⁻¹) pH 13.57 was added to the Mariotte flask traps; the NaOH solution upon reacting with CO₂ formed Na₂CO₃ and the released gases were a mixture of CH₄, H₂, N₂ and H₂S according to the technique described by Ley de Coss et al. (2018); the CO₂ trap was coupled to the biodigester using a Taygon hose (5 mm internal 35 cm length) to which a 31.8 mm 20 G hypodermic needle was attached. In all TG production evaluations, the result was corrected by difference to the gas production of the blank samples (200 mL of culture medium plus 20 mL of RL), as well as with the CH₄ concentration measurements using the gas trap technique with saline solution.

Statistical analysis. The experimental design was in a completely randomized design, total gas and CH₄ production data were analyzed with the PROC GLM procedure of the Statistical Analysis System (SAS) (SAS, 2011) and the means of the treatments were compared with Tukey’s test (P≤0.05).

RESULTS AND DISCUSSION

Chemical composition of the diets

Table 2 shows the chemical composition of the H. ruffa and L. leucocephala evaluated mixtures. By increasing the L. leucocephala proportion from 0.0% to 60% of the mixture, the NDF content decreased from 55.7% to 38.2%; FDA from 28.7% to 18.4%; while the CP content increased from 8.3% to 18.1% and the ethereal extract from 0.8% to 2.4%. The increase in CP content and reduction of NDF and FDA could have an impact on the production of TG and CH₄, during in vitro fermentation (Piñeiro et al., 2017). One of the factors that modify the fermentation pattern and products is the amount of fermentable NDF; in this study, the 100% H. ruffa fermentation, showed a higher production of total gases (Table 3). Grasses from tropical regions such as H. ruffa have a higher NDF amount than shrubs, which generates fermentation patterns with higher CH₄ production (Archimède et al., 2011).

Total gas production

Table 3 shows that increasing the L. leucocephala content in the mixture from 0.0% to 20% decreased the TG production at 24 and 96 h of incubation (P<0.05), at 48 and 72 h there was no difference between both treatments (P>0.05). When the L. leucocephala proportion increased to 40%, TG production was lower at all incubation hours (P<0.05) compared to the treatment without L. leucocephala, and there was also a difference (P<0.05) compared to the treatment where 20% L. leucocephala was included at all incubation hours.
Table 2. Chemical composition of *Hyparrhenia ruffa* and *Leucaena leucocephala* mixtures (g per 100 g of dry matter).

<table>
<thead>
<tr>
<th>Nutrient (%)</th>
<th>Proportion of <em>H. ruffa</em>: <em>L. leucocephala</em></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>100</td>
</tr>
<tr>
<td>Dry material</td>
<td>93.4</td>
</tr>
<tr>
<td>Ashes</td>
<td>9.0</td>
</tr>
<tr>
<td>Raw Fiber</td>
<td>28.8</td>
</tr>
<tr>
<td>NDF</td>
<td>55.7</td>
</tr>
<tr>
<td>ADF</td>
<td>28.7</td>
</tr>
<tr>
<td>Raw protein (N×6.25)</td>
<td>8.3</td>
</tr>
<tr>
<td>Ethereal extract</td>
<td>0.8</td>
</tr>
<tr>
<td>NFE</td>
<td>53.3</td>
</tr>
</tbody>
</table>

NDF, Neutral detergent fiber; ADF, Acid detergent fiber; N, Nitrogen; NFE, Nitrogen free extract.

When the *L. leucocephala* proportion of the mixture increased to 60%, there was no difference in total gas production (P > 0.05) compared to the treatment with 40% of *L. leucocephala*, indicating that with higher than 40% levels in the mixture, TG production did not increase. In all assessed treatments, the highest TG production was recorded between the 48th and 72nd hours of incubation.

When analyzing the proportion increase in the TG at 48 h, when the proportion of *L. leucocephala* increased from 0 to 20%, the production decreased by 1.25%, while when the proportion increased from 20% to 40%, the decrease was 41.42%, and when the proportion of *L. leucocephala* increased from 40% to 60%, the total gas production decreased by 13.98%.

In this research, the result indicates that the greatest effect in the TG decrease production is obtained when 40% of *L. leucocephala* is included. This effect could be related to the NDF and CP content of the different mixtures (Table 2), since, as the proportion of *L. leucocephala* in the mixture increases, the NDF content decreases and the CP content increases, due to a lower proportion of cell walls. With that, the fermentation pattern changes, the amount of potentially fermentable cell walls decreases, and consequently the amount and proportion of volatile fatty acids decreases. This decreases the proportion of acetate, H₂, CO₂ and increases the proportion of propionate. It is reported that in diets
with *L. leucocephala* containing 14% of CP, methane production is reduced (Lovett et al., 2004), and when NDF and FDA levels are lower, there is lower total gases production (Waghorn and Hegarty, 2011).

**CH₄ production**

A similar trend was observed in the CH₄ production to that obtained in TG production. The highest amount of CH₄ produced was in the treatment with 100% *H. ruffa* at 24 and 96 h of incubation (P<0.05). At 24 h of incubation, when the proportion of *L. leucocephala* increased from 0.0% to 20%, CH₄ production decreased (P<0.05), and the same response was observed when the *L. leucocephala* content was of 40 and 60%. At 48 and 72 h, there was a difference when 40 and 60% of *L. leucocephala* were included. The results obtained in this research show that increasing *L. leucocephala* content in the mixture decreases CH₄ production. Higher NDF and FDA content has been related to higher CH₄ production (Lopez et al., 2016; Vélez et al., 2018). When there is lower availability of digestible nutrients and higher cell wall content (NDF and FDA); CH₄ production is favored, while high levels of fast-fermenting carbohydrates and CP reduce CH₄ production (Owens et al., 1998).

**Table 4.** Cumulative CH₄ production in *in vitro* incubations (mL of gas per 20 g of fermented DM).

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Hyparrhenia ruffa: Leucaena leucocephala</th>
<th>Time (h)</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>24</td>
<td>48</td>
<td>72</td>
</tr>
<tr>
<td>100</td>
<td>111.97ab</td>
<td>176.19a</td>
<td>272.45a</td>
<td>451.81a</td>
</tr>
<tr>
<td>80:20</td>
<td>66.43b</td>
<td>173.09a</td>
<td>253.46a</td>
<td>354.99b</td>
</tr>
<tr>
<td>60:40</td>
<td>47.89b</td>
<td>74.68b</td>
<td>125.21b</td>
<td>167.47c</td>
</tr>
<tr>
<td>40:60</td>
<td>40.59 b</td>
<td>57.58b</td>
<td>114.22b</td>
<td>148.15c</td>
</tr>
<tr>
<td>SEM</td>
<td>10.41</td>
<td>16.25</td>
<td>14.34</td>
<td>21.43</td>
</tr>
<tr>
<td>MSD</td>
<td>27.22</td>
<td>42.47</td>
<td>37.49</td>
<td>40.32</td>
</tr>
</tbody>
</table>

SEM: Standard error of the mean; MSD: Minimum significant difference. a,b,c Means with different letters within the same column differ from each other, according to Tukey test (P<0.05).

**CONCLUSIONS**

By including higher than 20% levels of *L. leucocephala* in mixtures with *H. ruffa* grass in *in vitro* incubations, total gas production and CH₄ production decrease, due to the change in the fermentation pattern caused by decreasing the NDF and FDA content of the mixture, and at the same time increasing the crude protein content.

**REFERENCES**


